



## SPECIATION AND ANTIFUNGAL SUSCEPTIBILITY PROFILE OF CLINICAL CANDIDA ISOLATES

## Microbiology

Vijay Ambade\*

Associate professor, Department of Microbiology, B.R.L.A.B.V.M. G.M.C. Rajnandgaon, Chhattisgarh, India \*Corresponding Author

## ABSTRACT

**INTRODUCTION:** The epidemiological shift towards increase in prevalence of non-*albicans* *Candida* species, emergence of new *Candida* species and development of antifungal drug resistance to *Candida* species in global scenario made it relevant to identify *Candida* species with their antifungal susceptibility pattern. Hence, the present study was carried out.

**MATERIALS & METHODS:** Present prospective study was conducted at microbiology department of M.G.I.M.S. Sevagram, Maharashtra, India from January 2010 to December 2011. A total of 128 *Candida* strains, from various clinical specimens were identified by standard diagnostic techniques and with HiCrome *Candida* differential agar. All the identified *Candida* strains were tested for susceptibility against four antifungal agents amphotericin B, clotrimazole, ketoconazole and fluconazole by broth microdilution method (M27-A2) as per CLSI guidelines.

**RESULTS:** *C. albicans* (60.15%) were predominated over non-*albicans* *Candida* (39.85%). 3.90% isolates showed resistance to amphotericin-B, 7.03 % noted resistance to fluconazole while 3.90% isolates exhibited resistance to clotrimazole. *C. guilliermondii* found susceptible to all antifungal agents and none of *Candida* isolate found resistant to ketoconazole.

**CONCLUSION:** *Candida* speciation and antifungal resistance surveillance must be essential for studying the trend of antifungal resistance in a particular area and thereby to develop rational antifungal therapy.

## KEYWORDS

Candida species, antifungal susceptibility, broth microdilution method

## INTRODUCTION

*Candida* is unique among mycotic pathogens as it causes a broad spectrum of clinical manifestations ranging from mere mucocutaneous overgrowth to life threatening systemic infections<sup>(1)</sup>. Invasive *Candida* infections are one of major causes of morbidity and mortality in immunocompromised as well as critically ill immunocompetent patients<sup>(2)</sup>. Candidiasis has emerged itself as an alarming opportunistic disease due to increase in the number of immunocompromised, aged, receiving prolonged antibacterial and aggressive cancer chemotherapy or undergoing invasive surgical procedures and organ transplantation patients<sup>(3)</sup>. Though *C. albicans* is generally considered as the major pathogen, during last decades the epidemiological shift towards increase in the prevalence of non-*albicans* *Candida* species has been noted along with emergence of new *Candida* species<sup>(4)</sup>. The drug resistance scenario has been also increasing due to over growing use of random antifungal agents<sup>(4)</sup>. The commonly used antifungal drugs show significant variation in the susceptibility pattern among the different *Candida* species. Several previous studies reported the emergence of antifungal drug resistance *Candida* species in global scenario<sup>(5)</sup>. With this background, the present study was undertaken to speciate the *Candida* isolates and to know their antifungal susceptibility profile for using specific and sensitive drugs for better therapeutic outcome.

## MATERIALS AND METHODS

The present laboratory based prospective study was conducted at microbiology department of Mahatma Gandhi Institute of Medical Sciences, Sevagram, Maharashtra, India from January 2010 to

December 2011. A total of 128 *Candida* strains, from various clinical specimens were identified by standard diagnostic techniques (germ tube test, chlamyospore formation on corn meal agar, sugar fermentation, sugar assimilation tests and temperature tolerance at 45°C) and with HiCrome *Candida* differential agar (HiMedia, Mumbai, India). All the identified *Candida* strains were tested for susceptibility against four antifungal agents amphotericin-B (HIMEDIA), clotrimazole (SIGMA), ketoconazole (HIMEDIA) and fluconazole (HIMEDIA) by broth microdilution method (M27-A2) as per CLSI guidelines<sup>(6)</sup>. The results of antifungal susceptibility for fluconazole were interpreted as per CLSI interpretative criteria and for amphotericin B, ketoconazole as per Chakrabarti et al<sup>(7)</sup>. For clotrimazole, the results were interpreted as per Pelletier et al<sup>(8)</sup> and Martel et al<sup>(9)</sup>.

## RESULTS

Table 1: Different *Candida* species (n=128)

Candida Species	Number (%)
<i>Candida albicans</i>	77 (60.15)
<i>Candida tropicalis</i>	23 (17.96)
<i>Candida parapsilosis</i>	18 (14.06)
<i>Candida krusei</i>	09 (7.03)
<i>Candida guilliermondii</i>	01 (0.78)

Of the 128 *Candida* isolated, *C. albicans* (77/128; 60.15%) were predominated over non-*albicans* *Candida* (51/128; 39.84%). The non-*albicans* *Candida* species isolated were *C. tropicalis* (17.96%), *C. parapsilosis* (14.06%), *C. krusei* (7.03%) and *C. guilliermondii* (0.78%).

Table 2: *Candida* species susceptibility to antifungal agents

Candida species	Amphotericin B			Clotrimazole			Ketoconazole			Fluconazole		
	S	I	R	S	I	R	S	I	R	S	SD	R
<i>C. albicans</i> (77)	68 (88.31)	6 (7.79)	3 (3.89)	74 (96.10)	-	3 (3.89)	73 (94.80)	4 (5.19)	-	72 (93.50)	2 (2.59)	3 (3.89)
<i>C. tropicalis</i> (23)	19 (82.60)	3 (13.04)	1 (4.34)	22 (95.65)	-	1 (4.34)	21 (91.30)	2 (8.69)	-	21 (91.30)	1 (4.34)	1 (4.34)
<i>C. parapsilosis</i> (18)	18 (100)	-	-	17 (94.44)	-	1 (5.55)	13 (72.22)	5 (27.7)	-	18 (100)	-	-
<i>C. krusei</i> (9)	8 (88.88)	-	1 (11.11)	9 (100)	-	-	9 (100)	-	-	4 (44.44)	-	5 (55.55)
<i>C. guilliermondii</i> (1)	1 (100)	-	-	1 (100)	-	-	1 (100)	-	-	1 (100)	-	-
<b>Total (128)</b>	<b>114 (89.06)</b>	<b>9 (7.03)</b>	<b>5 (3.90)</b>	<b>123 (96.09)</b>	<b>-</b>	<b>5 (3.90)</b>	<b>117 (91.40)</b>	<b>11 (8.59)</b>	<b>-</b>	<b>116 (90.62)</b>	<b>3 (2.34)</b>	<b>9 (7.03)</b>

Among all *Candida* isolates tested, 3.90% *Candida* isolates showed resistance to amphotericin-B comprising *C. albicans* (3.89%), *C. tropicalis* (4.34%) and *C. krusei* (11.11%). Among the 3.90% *Candida*

isolates exhibited resistance to clotrimazole, 3.89% were *C. albicans*, 4.34% were *C. tropicalis* while 5.55% were *C. parapsilosis*. 7.03% *Candida* isolates noted resistance to fluconazole representing *C.*

*albicans* (3.89 %), *C. tropicalis* (4.34 %) and *C. krusei* (55.55 %). None of the *Candida* isolate was found to be resistant to ketoconazole. The isolate of *C. guilliermondii* was found to be susceptible to all the antifungal agents.

## DISCUSSION

Comparative study of different *Candida* species isolated in their studies by different workers<sup>(7,10,11,12,13,14)</sup> showed that *C. albicans* isolation was highest in each of them except Chakrabarti et al<sup>(8)</sup>, who reported *C. tropicalis* was highest (42 %) and *C. albicans* was 25 %. In our study also, the most frequently isolated species was *C. albicans* accounting for 60.15%. In the study of Baradkar et al<sup>(10)</sup>, *C. albicans* isolation rate was higher (70%) than our study while Pfeller et al<sup>(11)</sup>, Prasad et al<sup>(12)</sup>, Vijaya et al<sup>(13)</sup> and Grace et al<sup>(14)</sup> were reported 52%, 47.6%, 46% and 43.15% respectively which were lower than that our study. In the present study, most common non-*albicans Candida* species was *C. tropicalis* (17.96%) which approximates to that reported by Baradkar et al<sup>(10)</sup> (17.94%), but isolation rate lower than that showed by Vijaya et al<sup>(13)</sup> (35.29 %). Other non-*albicans Candida* species representing *C. parapsilosis* (14.06%), *C. krusei* (7.03%) and *C. guilliermondii* (0.78%) found in this study were also comparable with other workers<sup>(7,10,11,12,13,14)</sup>. In India, amphotericin-B is the drug of choice for invasive candidiasis with low or no resistance reports. However Chakrabarti et al<sup>(7)</sup> in his study reported the emergence of resistance to amphotericin-B in 15.4 % of *C. albicans* strains, 8.1 % in *C. tropicalis* strains and 33.3 % in *C. krusei* strains. In our study also, 3.9% *Candida* isolates showed resistance to amphotericin-B representing *C. albicans* (3.89 %), *C. tropicalis* (4.34 %) and *C. krusei* (11.11 %). Narang et al<sup>(15)</sup>, in a study of neonatal systemic candidiasis found 24% resistance against fluconazole. In our study, 6.25% *Candida* isolates showed resistance to fluconazole including *C. albicans* (1.56 %), *C. tropicalis* (4.34 %) and *C. krusei* (55.55 %). Narain et al<sup>(16)</sup> reported 18.75 % strains of *C. albicans*, 14.5% strains of *C. tropicalis* and all the strains of *C. krusei* resistant to fluconazole. In the study of Madhu Sharma et al<sup>(17)</sup>, resistance for fluconazole was found in *C. albicans* (14.28%), *C. tropicalis* (14.28%), *C. parapsilosis* (11.11%) and *C. krusei* (100%). None of the *Candida* isolate found resistant to ketoconazole in present study was exactly paralleled to findings of G. Sasikala et al<sup>(18)</sup>. However, Changdeo S. Aher<sup>(19)</sup> in his study reported 37.2 % *Candida* isolates resistant to ketoconazole while Khadka et al<sup>(20)</sup> reported it as 86%. 3.90% *Candida* isolates found resistant against clotrimazole in our study can be comparable with Khadka et al and Jayachandran AL et al<sup>(21)</sup> who reported it as 6% and 7.69% respectively. The resistant *Candida* isolates to clotrimazole found in our study were *C. albicans* (3.89 %), *C. tropicalis* (4.34 %) and *C. parapsilosis* (5.55%). Khadka et al noted 7.2% *C. albicans* while Jayachandran AL et al reported 7.52% *C. albicans*, 13.33% *C. tropicalis* and 9.09% *C. krusei* resistant against clotrimazole in their studies.

## CONCLUSION

In our study, *C. albicans* was predominant species and antifungal resistance to amphotericin-B, fluconazole & clotrimazole was also found less. However, *Candida* speciation and antifungal resistance surveillance must be essential for studying the trend of antifungal resistance in a particular area and thereby to develop rational antifungal therapy.

## REFERENCES

- Sardi JCO, Scorzoni L, Bernardi T, Fusco-Almeida AM, Mendes Giannini MJS. *Candida* species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. *J Med Microbiol*.2013;62(1):10-24.
- Eggimann P, Garbino J, Pittet D. Epidemiology of *Candida* species infections in critically ill non-immunosuppressed patients. *Lancet Infect Dis*. 2003;3(11):685-702.
- Jaya S, Harita V. *Candida* Species Isolated from Various Clinical Samples and Their Susceptibility Patterns to Antifungals. *J Med Microbiol/Infect Dis* 2013; 1:22-26.
- Yang YL, Cheng HH, Ho YA, Hsiao CF, Lo HJ. Fluconazole resistance rate of *Candida* species from different regions and hospital types in Taiwan. *J Microbiol/Immunol Infect*. 2003;36:187-91.
- White TC, Marr KA, Bowden RA. Clinical, cellular, and molecular factors that contribute to antifungal drug resistance. *ClinMicrobiol Rev*. 1998;11:382-402.
- National Committee for Clinical Laboratory Standards. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts—Second Edition: Approved Standard M27-A2. NCCLS, Wayne, PA, USA, 2002.
- Chakrabarti A, Ghosh A, Kanta A, Kumar P. In vitro antifungal susceptibility of *Candida*. *Ind J Med Res* 1995;102:13-9.
- Rene Pelletier, Joanne Peter, Cynthia Antin, Corina Gonzalez, Lauren Wood et al. Emergence of Resistance of *Candida albicans* to Clotrimazole in Human Immunodeficiency Virus-Infected Children: In Vitro and Clinical Correlations. *Journal of Clinical Microbiology*.2000;1563-1568.
- Claire M. Martel, Josie E. Parker, Oliver Bader, Michael Weig, Uwe Gross et al. Identification and Characterization of Four Azole-Resistant *erg3* Mutants of *Candida albicans*. *Antimicrobial Agents and Chemotherapy*.2010;4527-4533.
- Baradkar VP, Mathur M, Kumar S. Hichrom *Candida* agar for identification of *Candida* species. *Indian J Pathol/Microbiol* 2010;53:93-5
- Pfeller MA, Messer SA, Hollis RJ et al. Trends in species distribution and susceptibility

of fluconazole among blood stream isolates of *Candida* species in the United States. *DiagMicrobiol Infect Dis* 1999;33:4:311-7.

- Prasad KN, Agarwal J, Dixit AK, Tiwari DP, Dhole TN, Ayyageni A. Role of yeasts as nosocomial pathogens & their susceptibility to fluconazole & amphotericin B. *Indian J Med Res* 1999;110:11-17.
- Vijaya D, Nagarathnamma T. *Candida* Speciation Using Chrom Agar. *Journal of Clinical and Diagnostic Research*. 2011;5(4):755-757.
- Grace L, Tan, Ellena M Peterson. CHROM agar *Candida* medium for direct susceptibility testing of yeast from blood culture. *J ClinMicrobiol* 2005;43:4:1727-1731.
- Narang A, Agarwal PR, Chakrabarti A, Kumar P. Epidemiology of systemic candidiasis in a tertiary care neonatal unit. *J Trop paediatrics* 1998;44(2):104-108.
- S Narain, JS Shastri, M Mathur, PR Mehta. Neonatal systemic candidiasis in a tertiary care centre. *Indian Journal of Medical Microbiology*. 2003; 21 (1):56-58.
- Madhu Sharma, Sarita Yadav, Parma and Uma Chaudhary. *Candida* blood stream infections in neonates. *Ijpbs* 2011; 2(2).
- Sasikala G, Udayasri B. Speciation and antifungal susceptibility profiles of *Candida* isolates from vaginitis patients attending STD Clinic at a Tertiary Care Hospital. *J NTR Univ Health Sci* 2018;7:94-7.
- Changdeo S. Aher. Species distribution, virulence factors and antifungal susceptibility profile of *Candida* isolated from Oropharyngeal lesions of HIV infected patients. *Int.J.Curr.Microbiol.App.Sci* 2014 3(1):453-460.
- Sundar Khadka, Jeevan Bahadur Sherchand, Bharat Mani Pokhrel et al. Isolation, speciation and antifungal susceptibility testing of *Candida* isolates from various clinical specimens at a tertiary care hospital, Nepal. *BMC Res Notes* 2017;10:218.
- Jayachandran AL, Radhika Katragadda, Thyagarajan Ravinder et al. Antifungal susceptibility pattern among *Candida* species: An Evaluation of Disc Diffusion and Broth Micro-dilution Method *J Microbiol Infect Dis* 2018;8 (3).